

MTN 026 Laboratory Training Follow Up Visits

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Objectives



- ◆ Overview of Lab testing
- ◆ Collection of Pelvic Samples
- ◆ Collection of Anorectal Specimens
- ◆ Specimen Management
- ◆ Q&A

Overview of Lab Testing by Visit

	VST 1 SCR	VST 2 ENR	VST 3 Dose	VST 4-6	VST 7-12	VST 13 Final Dose	VST 14-16
UA and Culture	★	★	★	★	★	★	★
Urine GC/CT	X	★	★	★	★	★	★
Urine hCG (♀)	X	X			7		14
CBC/diff/plt	X	★	★	★	★	★	★
AST/ALT/Creatinine	X	★	★	★	★	★	16
HIV-1 and HIV-2 serology	X	X			7		16
Syphilis Serology	X	★	★	★	★	★	★
HSV-1, HSV-2, HBsAg, and HCV Serologies	X						
Coagulation (PT/INR)	X						
Plasma Archive/Storage		X			7		16
Plasma PK (Vsts 3 and 13: pre and 30-60 or 120 mins)			◆	X	7 and 8	◆	X

★ As indicated, ◆ Randomized assigned time points

Overview of Lab Testing by Visit

	VST 1 SCR	VST 2 ENR	VST 3 Dose	VST 4-6	VST 7-12	VST 13 Final Dose	VST 14-16
Vaginal NAAT for GC/CT (♀)	X						
CVL for PD/PK (♀)		X		♦		X	♦
CVF and Biopsies for PK (♀)				♦		X	♦
Pap Test (♀)	★						
Rectal HSV-1/2 detection	★	★	★	★	★	★	★
Rectal NAAT for GC/CT	X	★	★	★	★	★	★
Rectal Fluid for Microflora		X	♦			♦	
Rectal Sponge Mucosal Safety		X	♦			♦	
Rectal Fluid and Biopsies for PK (Vsts 3 and 13: 30-60 or 120 mins)			♦	♦	7 and 8	♦	♦
Rectal Enema for PD/PK		X		♦			♦
Rectal Biopsies for PD (Vsts 3 and 13: 30-60 or 120 mins)		X	♦	♦		♦	♦
Rectal Biopsies for GE, Histology, T Cell Pheno, and Proteomics		X	♦			♦	

★ As indicated, ♦ Randomized assigned time points



Urine Specimens

- ◆ Urine hCG (♀) (Visits 7 and 14)
- ◆ Urinalysis and culture (if indicated)
- ◆ NAAT for GC/CT (if indicated)

Blood Specimens



- 💧 HIV Testing (Visits 7 and 16)

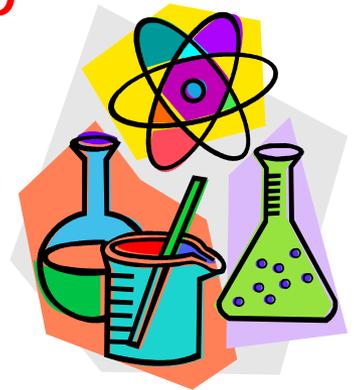
- 💧 In the case of seroconverters, notify MTN LC immediately by submitting a MTN LC HIV Testing Notification and Query Form to mtnvirology@mtnstopshiv.org

- 💧 Plasma for storage (Visits 7 and 16)

- 💧 Chemistries (Visit 16)

- 💧 Syphilis Serology (if indicated)

- 💧 CBC with diff and platelets (if indicated)



MTN Laboratory Center HIV Testing Notification & Query Form

HIV Testing Notification

www.mtnstopshiv.org



Study	
PTID	
Site/Contact Person	
Last Update	

Form Closed

Please check one:

- Notification (LC response not required)
- Query (Waiting for LC Response)

Please copy/paste this table for test results from additional visits.

QUERY DATE:					
	Screening Immunoassay 1	Screening Immunoassay 2	Confirmatory Assay	HIV RNA	Other Test
Visit Code					
Testing Date					
Kit Name					
Kit Lot/Exp (optional)					
Result					

SITE COMMENTS/QUERY:

LC RESPONSE:

Participant Final Outcome

- HIV NEGATIVE
- HIV POSITIVE
- OTHER (Please describe further)

Blood Specimens

💧 Plasma for PK

- 💧 Collect 10mL EDTA blood at each time point.
- 💧 For visits 3 and 13, a pre-dose draw along with a randomized time point at 30-60 mins or 120 mins after dosing will be obtained. Be sure to identify time point by circling it on the LDMS TS.

<input type="checkbox"/>	Blood – <i>Plasma PK Post-dose (BLD)</i> Collection Time: ____:____ Hour : Min	EDT (purple top)	PL1	N/A	Store in 1.0 ml aliquots and freeze within 8 hours of collection. Enter PK into Other Spec ID field of LDMS. PK Time Point (circle one): 30-60 min or 120 min
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- 💧 At visits 4-8 and 14-16 only one time point is required.



Pelvic Samples (♀)

(Visits r4-6, 13, r14-16)

- ◇ CVF swab for PK
 - ◇ Collect within **one hour** of PK blood draw.
 - ◇ Weigh dacron swab/collection kit before and after collection using same analytical scale measuring to 0.1mg. Record weights onto LDMS TS.
 - ◇ Swab may be pre-cut and held with forceps or cut post collection. For details see SSP.
 - ◇ Keep refrigerated and freeze within 2 hours.

- ◇ Cervicovaginal Lavage for PD/PK

Pelvic Samples (♀)

(Visits r4-6, 13, r14-16)

- ◇ Cervical biopsies for PK
 - ◇ Weigh two (2) labeled cryovial using an analytical scale with a sensitivity of 0.1mg. Document the pre-weight on the LDMS TS.
 - ◇ Collect two (2) biopsies and place one in bottom of each pre-weighed cryovial.
 - ◇ Obtain the post-weight for each cryovial containing a biopsy using the same analytical scale and document on the LDMS TS.
 - ◇ Immediately freeze in liquid nitrogen or a dry ice alcohol bath. Document frozen time.
 - ◇ Store frozen biopsies at $\leq -70^{\circ}\text{C}$.



Rectal Specimens

- ❖ Rectal Swabs
 - ❖ Microflora (Visits 3 and 13)
 - ❖ PK (Visits 3, r4-6, 7, 8, 13, and r14-16)
 - ❖ NAAT for GC/CT (if indicated)
 - ❖ HSV-1 and -2 (if indicated)
- ❖ Rectal Sponge for M Safety (Visits 3 and 13)
- ❖ Rectal Enema for PD (Visits r4-6 and r14-16)
- ❖ Rectal Biopsies in order of importance
 - ❖ PK (Visits 3, r4-6, 7, 8, 13, and r14-16)
 - ❖ Gene Expression (Visits 3 and 13)
 - ❖ Histology (Visits 3 and 13)
 - ❖ PD (Visits 3, r4-6, 13, and r14-16)
 - ❖ T Cell Phenotyping (Visits 3 and 13)
 - ❖ Proteomics (Visits 3 and 13)



Collection of Rectal Specimens

- ❖ Rectal swab for PK
 - ❖ Collect within 1 hour of PK blood draw
 - ❖ Weigh dacron swab/collection kit before and after collection using same analytical scale measuring to 0.1mg. Record weights onto LDMS TS.
 - ❖ Swab may be pre-cut and held with forceps or cut post collection. For details see SSP.
 - ❖ Keep refrigerated and freeze within 2 hours.



Collection of Rectal Specimens

❖ Rectal Biopsies for PK

- ❖ Weigh five (5) labeled cryovial using an analytical scale with a sensitivity of 0.1 mg. Document the pre-weight on the LDMS TS.
- ❖ Collect five (5) biopsies and place one in bottom of each pre-weighed cryovial.
- ❖ Obtain the post-weight for each cryovial containing a biopsy using the same analytical scale and document on the LDMS TS.
- ❖ Immediately freeze in liquid nitrogen or a dry ice alcohol bath. Document frozen time.
- ❖ Store frozen biopsies at $\leq -70^{\circ}\text{C}$.



Specimen Management

- ◆ All specimens must be tracked according to site Chain of Custody.
- ◆ CRFs are required for specimens reported to SCHARP.
- ◆ Specimens to be shipped to the Laboratory Center must be accompanied by an LDMS tracking sheet and entered into LDMS.

Any Questions?

